

blood

2013 121: 4032-4035
Prepublished online April 5, 2013;
doi:10.1182/blood-2012-12-453076

The laboratory and the direct oral anticoagulants

Armando Tripodi

Updated information and services can be found at:

<http://bloodjournal.hematologylibrary.org/content/121/20/4032.full.html>

Articles on similar topics can be found in the following Blood collections

[Blood Spotlight](#) (4 articles)

[Thrombosis and Hemostasis](#) (579 articles)

Information about reproducing this article in parts or in its entirety may be found online at:

http://bloodjournal.hematologylibrary.org/site/misc/rights.xhtml#repub_requests

Information about ordering reprints may be found online at:

<http://bloodjournal.hematologylibrary.org/site/misc/rights.xhtml#reprints>

Information about subscriptions and ASH membership may be found online at:

<http://bloodjournal.hematologylibrary.org/site/subscriptions/index.xhtml>

Blood (print ISSN 0006-4971, online ISSN 1528-0020), is published weekly by the American Society of Hematology, 2021 L St, NW, Suite 900, Washington DC 20036.
[Copyright 2011 by The American Society of Hematology; all rights reserved.](#)



The laboratory and the direct oral anticoagulants

Armando Tripodi

Angelo Bianchi Bonomi Hemophilia and Thrombosis Center, Department of Clinical Sciences and Community Health, Università degli Studi di Milano and Istituto di Ricerca e Cura a Carattere Scientifico Cà Granda Ospedale Maggiore Foundation, Milano, Italy

Although direct oral anticoagulants do not need laboratory testing for dose adjustment, there are instances when laboratory measurement of the drug anticoagulant effect may be useful. They include before initiation of treatment, before surgical or invasive procedures, on the occasion of hemorrhagic or thrombotic events, and whenever immediate reversal of anticoagulation is needed. Choice of tests should be primarily based on their prompt availability. Accordingly, the dilute-thrombin or the ecarin clotting times are best suited for dabigatran and the prothrombin time or the anti-FXa for rivaroxaban. (*Blood*. 2013;121(20):4032-4035)

The direct oral anticoagulants (DOAC), which include the thrombin inhibitor dabigatran and the anti-Xa agents rivaroxaban, apixaban, and edoxaban, are in clinical use in many countries (Figure 1). Clinical trials have shown that DOAC, unlike vitamin K antagonists (VKA), are effective and safe when administered without dose adjustment based on laboratory testing.¹ However, it is a misconception to believe that the laboratory will not have a role in the management of patients treated with DOAC. This review is aimed at discussing:

- The situations where the laboratory may help clinicians with decision making in the management of treated patients;
- The most appropriate tests to be used; and
- The interpretation of results.

Need for testing

Although the experience is still limited, one may identify situations when the laboratory measurement for DOAC is potentially useful and others that would require further evaluation (Table 1).

Potentially useful

At baseline. Laboratory (and clinical) evaluation is useful for excluding an underlying bleeding tendency or renal insufficiency and to determine if the test(s) that will be used for measuring the drug effect is (are) normal at baseline. In addition to the prothrombin time (PT) and partial thromboplastin time (APTT), other tests to be included are blood cell counts and others that will depend on which drug is used for treatment (see below). Assessment of creatinine clearance is of paramount importance as DOAC are largely excreted via the kidney.¹ Therefore, even mild renal insufficiency may lead to drug accumulation during treatment, thus increasing the bleeding risk. Assessment of creatinine clearance is indicated not only before starting treatment but also at regular intervals during treatment, as kidney function may deteriorate rapidly, especially in the elderly.

Preoperatively. Owing to their short half-life, the effect of DOAC may be rapidly dissipated by discontinuation of the treatment before surgical or invasive procedures. However, there may be patients with mild, perhaps unrecognized renal insufficiency in whom the clearance may be delayed.² Preoperative laboratory testing of the drug anticoagulant effect may therefore help to prevent the risk of

life-threatening hemorrhage^{2,3} and expensive therapeutic interventions aimed at reversing anticoagulation.²

Adverse events. Patients referred to emergency departments with thrombosis or hemorrhage should be tested to see whether they are under- or overanticoagulated. Laboratory testing may also help in the event of life-threatening (intracranial) hemorrhage to help decide whether immediate anticoagulation reversal is needed and to judge subsequently whether neutralization is achieved. There are not yet antidotes for DOAC, and current recommendations for reversing their effects are based on the experience accumulated with traditional anticoagulants (ie, infusion of prothrombin complex concentrates [PCC] or recombinant FVIIa). Although the experience with these procoagulants in the reversal of DOAC effects is still limited, a recent study⁴ suggested a role for laboratory testing in assessing drug neutralization. Healthy subjects were treated with dabigatran or rivaroxaban for 2.5 days before reversal of anticoagulation by PCC or saline infusion. Laboratory testing showed that PCC were able to neutralize the effect of rivaroxaban but not that of dabigatran.⁴ It is still unknown whether these results reflect what occurs in vivo and to what extent laboratory testing may help to judge reversal. However, if different drugs will have different antidotes and/or reversal strategies, physicians in emergency departments need to be able to assess the levels of anticoagulation and the specific DOAC being used by patients. Simple laboratory detection (eg, in the urine) of the type of drug⁵ might be useful.

Others

Coagulation tests might be considered upon attainment of stable anticoagulation (1-2 weeks after initiation) to provide the level of the anticoagulation achieved chronically. This information might be useful to interpret subsequent results.

Drug anticoagulant levels could be measured occasionally at the time of medical visits to assess adherence to treatment. However, it should be realized that given the short DOAC half-life (8-15 hours), a dose missed a few days earlier than testing might not be detected in the laboratory.

Since DOAC may interact with other drugs,⁶ comparison of their anticoagulant effects measured before and after the introduction of additional drugs may help to confirm the degree of known drug-to-drug interactions or to unravel unknown interactions.

DOAC are prescribed at fixed dosage, so individuals with extreme body weights may benefit from dose adjustment to avoid

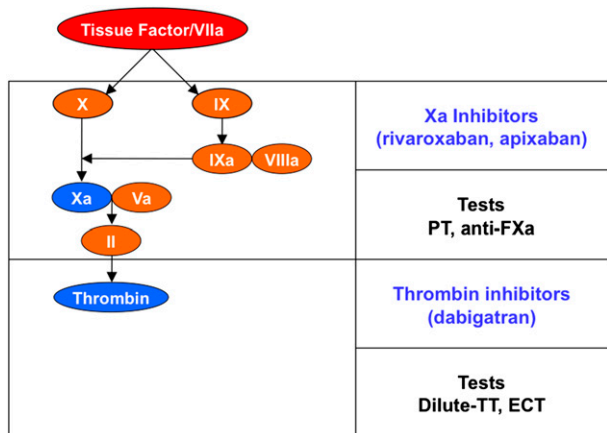


Figure 1. Schematic representation of coagulation. DOACs and tests suggested for the measurement of their anticoagulant effect. ECT, ecarin clotting time; PT, prothrombin time; TT, thrombin clotting time.

under- or overanticoagulation. This issue was not evaluated in phase 3 trials and warrants assessment when DOAC will be used in real life. Information based on laboratory testing and clinical events recorded during treatment could help when making decisions about these patients.

Which test(s) for which drug

The following sections are aimed at discussing the choice of tests for the currently available drugs. The choice is based on the limited experience and literature data accumulated to date and on the following practical considerations:

- Prompt test availability;
- Linearity and adequacy of test response to increasing dosage; and
- Amenability to standardization (ie, comparability of results between laboratories using the same method but different reagents). Standardization is mandatory for patients on VKA, but it is also important for DOAC. Assuming that future clinical studies will determine specific cutoff values able to alert clinicians to bleeding risk, these values could be generalized to all laboratories, regardless of the reagent used for testing, only if the tests are standardized.

Dabigatran

Dabigatran prolongs the clotting times of tests based on thrombin generation and fibrinogen-to-fibrin conversion. Among them, the most promising are the thrombin time (TT) and ecarin clotting time (ECT).⁷ TT reflects fibrinogen-to-fibrin conversion and is performed by measuring plasma clotting times upon addition of thrombin. TT is

Table 1. Situations when the measurement of the DOAC anticoagulant effect is potentially useful and others that would require consideration

Potentially useful	Others
At baseline (before initiation of treatment)	When chronic anticoagulation is achieved (1-2 weeks after initiation)
Before surgical or invasive procedures	During clinical visits
During adverse events (hemorrhage or thrombosis)	Soon before and after introducing additional drugs
Need for immediate reversal of anticoagulation	Low or high body weight

Table 2. Expected plasma concentrations of dabigatran or rivaroxaban after treatment

Drug	C _{peak} (range)	C _{trough} (range)
Dabigatran*	175 ng/mL (117-275 ng/mL)	91 ng/mL (61-143 ng/mL)
Rivaroxaban†	215 ng/mL (22-535 ng/mL)	32 ng/mL (6-239 ng/mL)

*Steady-state geometric mean dabigatran plasma concentration (25th-75th percentile range) measured around 2 hours (C_{peak}) or 12 hours (C_{trough}) after 150 mg dabigatran administration twice daily.¹⁰ Dilute thrombin clotting time or ECT at C_{peak} are approximately prolonged 2 or 3 times the baseline value, respectively.⁸ Between-reagent variability should be considered when interpreting results.

†Steady-state geometric mean rivaroxaban plasma concentration (90% prediction interval) measured around 2 hours (C_{peak}) or 24 hours (C_{trough}) after 20 mg rivaroxaban administration once daily.¹⁷ PT at C_{peak} is approximately prolonged 1.5 times the baseline value.^{16,19} Between-reagent variability should be considered when interpreting results.^{16,19,26,30}

readily available in most laboratories; the clotting time prolongation is linearly and dose-dependently related to dabigatran concentrations.⁸ However, responsiveness to increasing dosage is excessive: TT prolongations induced by the typical 175 ng/mL dabigatran plasma concentration (observed in patients taking 150 mg twice daily)^{9,10} may be 10 times the baseline value.⁸ Hence, a normal TT should rule out a dabigatran anticoagulant effect, but the degree of prolongation poorly reflects drug concentration. Recently, a dilute TT adequately responsive to dabigatran (ie, prolongation up to 2 times the baseline value at 175 ng/mL dabigatran) (Table 2) has become available and may be used for dabigatran measurement.^{11,12}

Ecarin is a commercially available snake venom that converts FII into meizothrombin. The dabigatran-induced inhibition of meizothrombin is measured by synthetic substrates or clotting assays.¹³ ECT proved to be linearly and dose-dependently related to dabigatran concentrations: ECT prolongations induced by 175 ng/mL amount to 3 times the baseline value⁸ (Table 2). Presently, ECT is not readily available in most laboratories but could be easily implemented in many coagulation platforms.^{13,14}

The APTT is another test that can be used for dabigatran; the APTT prolongation in response to 175 ng/mL dabigatran amounts to 1.7 times the baseline value but is not linearly related to the concentration.⁸ Furthermore, the between-reagent variability associated with the APTT makes the interpretation of results obtained with one reagent not necessarily applicable to others. The PT is relatively insensitive to dabigatran⁸ and should not be used for this drug.

In summary, the dilute-TT or ECT are the tests of choice for dabigatran. The first is readily available even in an emergency and can be easily implemented in all coagulometers. The second (although not yet widely available) could become available in clinical laboratories as the ecarin venom is commercially available. Both dilute-TT and ECT are fairly responsive to dabigatran⁸ and might be easier to standardize than the APTT.

Rivaroxaban

Rivaroxaban inhibits FXa and prolongs the clotting times of tests assessing coagulation downstream from FXa. Among them, the most promising are the anti-FXa and PT tests.⁷

Anti-FXa is based on the measurement of residual FXa with synthetic substrates upon mixing plasma with FXa. This test, presently used for low-molecular-weight heparins, has been modified to be fairly responsive to rivaroxaban.¹⁵

PT prolongations are adequate to detect rivaroxaban activity within the range of concentrations observed in treated patients.¹⁶ On average, PT is prolonged 1.5 times the baseline value at 215 ng/mL rivaroxaban¹⁶ (ie, the plasma concentration observed in patients

taking 20 mg once daily)¹⁷ (Table 2). Hence, prolongations are greater in patients who have received an overdose of the drug.

In summary, the PT or the anti-FXa assay can be used for patients on rivaroxaban. However, the anti-FXa is not readily available in most laboratories, especially at night. Moreover, standardization as assessed in a collaborative survey is poor: the average interlaboratory variation of results obtained while measuring the same rivaroxaban-spiked plasmas by means of local methods was high (ie, 25%).¹⁸ Conversely, the PT is readily available even in an emergency and its standardization could be obtained as described^{19,20} by expressing results as PT ratio (patient-to-normal) corrected by means of a sensitivity index. Such an index can be determined for working thromboplastins relative to an international standard. The international normalized ratio (used for VKA) should not be used for rivaroxaban because it increases the drug-induced between-thromboplastin variability.¹⁹

Apixaban

Limited experience is available for apixaban and laboratory testing. While the PT is in general poorly responsive, the anti-FXa assay proved to be highly correlated and adequately responsive to the apixaban plasma concentrations.²¹ The problems of availability and standardization of the anti-FXa assay discussed above for rivaroxaban also apply to apixaban.

Timing of testing

Onset/offset actions of DOAC are relatively fast. Peak plasma levels are reached 2 hours after ingestion of dabigatran^{8,10} or rivaroxaban.¹⁷ Rivaroxaban plasma concentration (in patients taking 20 mg once daily) is as low as 32 ng/mL¹⁷ at 24 hours and dabigatran plasma concentration (in patients taking 150 mg twice daily) is reduced to 91 ng/mL after 12 hours¹⁰ (Table 2). Consequently, the interpretation of test results is heavily dependent on the timing of blood sampling. It is still unclear whether the peak or trough drug concentration should be evaluated when the choice of timing for blood drawing is feasible.

Expression of results

Results of coagulation tests (ie, dilute-TT, ECT, and PT) can be expressed as clotting time ratio (patient-to-normal). Alternatively, clotting times can be converted into drug concentration equivalent by interpolation of patient clotting times from a dose-response curve, constructed by plotting clotting times for calibration plasmas spiked with known amounts of drugs versus their target concentrations. This result expression has been proposed for dilute-TT¹² and anti-FXa assays.¹⁸ Whether clotting time prolongation or drug concentration equivalent is the preferred way of expressing results has not yet been established.

Alert values

Test values able to alert clinicians to bleeding or thrombotic risk have not yet been established because the experience to date is based mainly on normal plasma spiked in vitro with increasing amounts of each DOAC. Accurate determination of alert values requires clinical observations in treated patients combined with standardized laboratory testing and will be undertaken when DOAC are more widely used.

DOAC effect on common hemostatic parameters

Owing to their mode of action, it is anticipated that DOAC affect the measurement of some of the most common hemostatic parameters. Here some examples:

- Antithrombin activity is overestimated in patients on rivaroxaban or dabigatran when the target enzyme used for testing is FXa or thrombin, respectively.^{22,23} This can be avoided by using FXa or thrombin as a target enzyme when the drug used for treatment is dabigatran or rivaroxaban, respectively.
- Fibrinogen, when measured as clotting activity, is underestimated in patients on dabigatran.²³ The degree of underestimation is reagent dependent and is likely due to different types or concentrations of thrombin used for testing.
- Tests for activated protein C resistance may be affected by DOAC when testing is based on paired APTT with or without activated protein C.^{23,24} Because of this effect, plasmas from heterozygous FV Leiden carriers might resemble wild-types when patients are treated with DOAC.^{23,24}
- FXIII when measured by chromogenic substrates may be underestimated in patients on dabigatran,²⁵ simply because FXIII is activated by thrombin.
- Although few data are available, it is possible that measurements of proteins C and S, performed with clotting techniques,²⁶ individual coagulation factors,^{26,27} and the search for lupus anticoagulants,²⁸ may also be affected by DOAC.

All of the above considerations point to the conclusion that caution should be exerted in the interpretation of results of hemostatic parameters measured for patients on DOAC.

Conclusions

Although DOAC do not require laboratory testing for dose adjustment, there are occasions when laboratory investigations may be useful. Many tests can be used, and the choice should be primarily based on their prompt availability. In the future, there will be millions of patients worldwide treated with DOAC. Some of them may bleed anywhere and anytime. This situation requires the availability of simple assays that can be run in both large and small hospitals. To this end, the dilute-TT or ECT tests are best suited for dabigatran and the PT or anti-FXa for rivaroxaban. The choice between TT and ECT or between PT and anti-FXa depends on clinical observations that are still lacking. Owing to the rather large between-reagent variability, laboratories should define their own reagent responsiveness to DOAC.²⁹ Results should be interpreted with caution if responsiveness is unknown.³⁰

Authorship

Contribution: A.T. prepared the manuscript, and no other individuals were involved in the writing or editing of the manuscript.

Conflict-of-interest disclosure: A.T. received speaker's fees on the occasion of educational meetings organized by Instrumentation Laboratory.

Correspondence: A. Tripodi, Via Pace 9, 20122-Milano, Italy; e-mail: armando.tripodi@unimi.it.

References

1. Tripodi A, Palareti G. New anticoagulant drugs for treatment of venous thromboembolism and stroke prevention in atrial fibrillation. *J Intern Med*. 2012; 271(6):554-565.
2. Warkentin TE, Margetts P, Connolly SJ, Lamy A, Ricci C, Eikelboom JW. Recombinant factor VIIa (rFVIIa) and hemodialysis to manage massive dabigatran-associated postcardiac surgery bleeding. *Blood*. 2012;119(9):2172-2174.
3. Harper P, Young L, Merriman E. Bleeding risk with dabigatran in the frail elderly. *N Engl J Med*. 2012; 366(9):864-866.
4. Eerenberg ES, Kamphuisen PW, Sijpkens MK, Meijers JC, Buller HR, Levi M. Reversal of rivaroxaban and dabigatran by prothrombin complex concentrate: a randomized, placebo-controlled, crossover study in healthy subjects. *Circulation*. 2011;124(14):1573-1579.
5. Harenberg J, Kraemer R. Measurement of the new anticoagulants. *Thromb Res*. 2012; 129(suppl 1):S106-S113.
6. Pengo V, Crippa L, Falanga A, et al; Italian Federation of Thrombosis Centers. Questions and answers on the use of dabigatran and perspectives on the use of other new oral anticoagulants in patients with atrial fibrillation. A consensus document of the Italian Federation of Thrombosis Centers (FCSA) *Thromb Haemost*. 2011;106(5):868-876.
7. Baglin T, Hillarp A, Tripodi A, Elalamy I, Buller H, Ageno W. Measuring oral direct inhibitors (ODIs) of thrombin and factor Xa: a recommendation from the Subcommittee on Control of Anticoagulation of the Scientific and Standardisation Committee of the International Society on Thrombosis and Haemostasis. *J Thromb Haemost*. 2013.
8. Stangier J, Rathgen K, Stähle H, Gansser D, Roth W. The pharmacokinetics, pharmacodynamics and tolerability of dabigatran etexilate, a new oral direct thrombin inhibitor, in healthy male subjects. *Br J Clin Pharmacol*. 2007;64(3):292-303.
9. Douxfils J, Mullier F, Robert S, Chatelain C, Chatelain B, Dogné JM. Impact of dabigatran on a large panel of routine or specific coagulation assays. Laboratory recommendations for monitoring of dabigatran etexilate. *Thromb Haemost*. 2012;107(5):985-997.
10. European Medicine Agency. Annex I. Dabigatran summary of product characteristics. Available at: <http://www.ema.europa.eu/>. Accessed February 2013.
11. Avecilla ST, Ferrell C, Chandler WL, Reyes M. Plasma-diluted thrombin time to measure dabigatran concentrations during dabigatran etexilate therapy. *Am J Clin Pathol*. 2012;137(4): 572-574.
12. Stangier J, Feuring M. Using the HEMOCLOT direct thrombin inhibitor assay to determine plasma concentrations of dabigatran. *Blood Coagul Fibrinolysis*. 2012;23(2):138-143.
13. Nowak G. The ecarin clotting time, a universal method to quantify direct thrombin inhibitors. *Pathophysiol Haemost Thromb*. 2003;33(4): 173-183.
14. Siegmund R, Boer K, Poeschel K, Wolf G, Deufel T, Kiehnopf M. Comparison of the ecarin chromogenic assay and different aPTT assays for the measurement of argatroban concentrations in plasma from healthy individuals and from coagulation factor deficient patients. *Thromb Res*. 2008;123(1):159-165.
15. Samama MM, Amiral J, Guinet C, Perzborn E, Depasse F. An optimised, rapid chromogenic assay, specific for measuring direct factor Xa inhibitors (rivaroxaban) in plasma. *Thromb Haemost*. 2010;104(5):1078-1079.
16. Samama MM, Martinoli JL, LeFlem L, Guinet C, Plu-Bureau G, Depasse F, Perzborn E. Assessment of laboratory assays to measure rivaroxaban—an oral, direct factor Xa inhibitor. *Thromb Haemost*. 2010;103(4):815-825.
17. European Medicine Agency. Annex I. Rivaroxaban summary of product characteristics. Available at: <http://www.ema.europa.eu/>. Accessed February 2013.
18. Samama MM, Contant G, Spiro TE, et al; Rivaroxaban Anti-Factor Xa Chromogenic Assay Field Trial Laboratories. Evaluation of the anti-factor Xa chromogenic assay for the measurement of rivaroxaban plasma concentrations using calibrators and controls. *Thromb Haemost*. 2012;107(2):379-387.
19. Tripodi A, Chantarangkul V, Guinet C, Samama MM. The international normalized ratio calibrated for rivaroxaban has the potential to normalize prothrombin time results for rivaroxaban-treated patients: results of an in vitro study. *J Thromb Haemost*. 2011;9(1):226-228.
20. Harenberg J, Marx S, Krämer R, Giese C, Weiss C. Determination of an international sensitivity index of thromboplastin reagents using a WHO thromboplastin as calibrator for plasma spiked with rivaroxaban. *Blood Coagul Fibrinolysis*. 2011; 22(8):637-641.
21. Becker RC, Yang H, Barrett Y, Mohan P, Wang J, Wallentin L, Alexander JH. Chromogenic laboratory assays to measure the factor Xa-inhibiting properties of apixaban—an oral, direct and selective factor Xa inhibitor. *J Thromb Thrombolysis*. 2011;32(2):183-187.
22. Mani H, Hesse C, Stratmann G, Lindhoff-Last E. Rivaroxaban differentially influences ex vivo global coagulation assays based on the administration time. *Thromb Haemost*. 2011; 106(1):156-164.
23. Lindahl TL, Baghaei F, Blixter IF, et al; Expert Group on Coagulation of the External Quality Assurance in Laboratory Medicine in Sweden. Effects of the oral, direct thrombin inhibitor dabigatran on five common coagulation assays. *Thromb Haemost*. 2011;105(2):371-378.
24. Hillarp A, Baghaei F, Fagerberg Blixter I, et al. Effects of the oral, direct factor Xa inhibitor rivaroxaban on commonly used coagulation assays. *J Thromb Haemost*. 2011;9(1):133-139.
25. Halbmayr WM, Weigel G, Quehenberger P, et al. Interference of the new oral anticoagulant dabigatran with frequently used coagulation tests. *Clin Chem Lab Med*. 2012;50(9):1601-1605.
26. Douxfils J, Mullier F, Loosen C, Chatelain C, Chatelain B, Dogné JM. Assessment of the impact of rivaroxaban on coagulation assays: laboratory recommendations for the monitoring of rivaroxaban and review of the literature. *Thromb Res*. 2012;130(6):956-966.
27. Asmis LM, Alberio L, Angelillo-Scherrer A, et al. Rivaroxaban: Quantification by anti-FXa assay and influence on coagulation tests: a study in 9 Swiss laboratories. *Thromb Res*. 2012;129(4): 492-498.
28. van Os GM, de Laat B, Kamphuisen PW, Meijers JC, de Groot PG. Detection of lupus anticoagulant in the presence of rivaroxaban using Taipan snake venom time. *J Thromb Haemost*. 2011; 9(8):1657-1659.
29. Freyburger G, Macouillard G, Labrousse S, Sztark F. Coagulation parameters in patients receiving dabigatran etexilate or rivaroxaban: two observational studies in patients undergoing total hip or total knee replacement. *Thromb Res*. 2011; 127(5):457-465.
30. van Veen JJ, Smith J, Kitchen S, Makris M. Normal prothrombin time in the presence of therapeutic levels of rivaroxaban. *Br J Haematol*. 2013;160(6):859-861.